Detecting toxic *Microcystis* in the lower Great Lakes

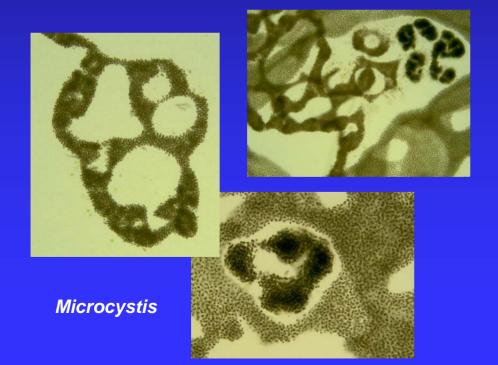


Microcystis in the Great Lakes

- Colonial cyanobacterial HAB
- Forms blooms and scums
 - Taste/odor issues in drinking water
 - Loss of recreational and fishing value to affected waters

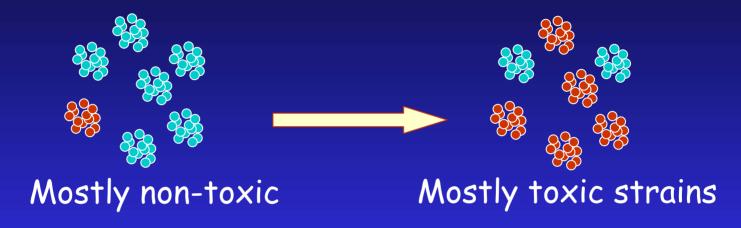
Put-In-Bay, Lake Erie

Blooms can be toxic or non-toxic

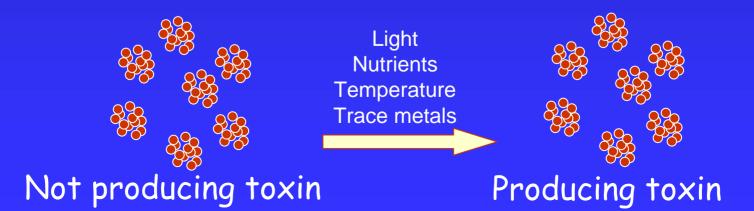


What makes a cyanobacterial bloom toxic?

Shift in community composition

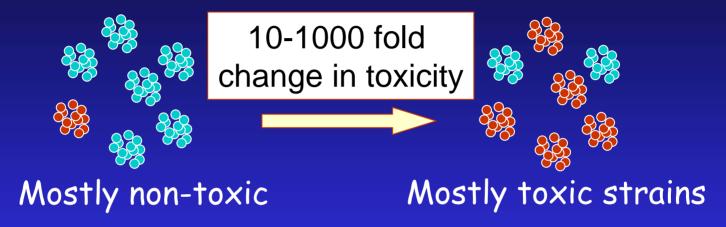


Stimulation of toxin production by environmental factors

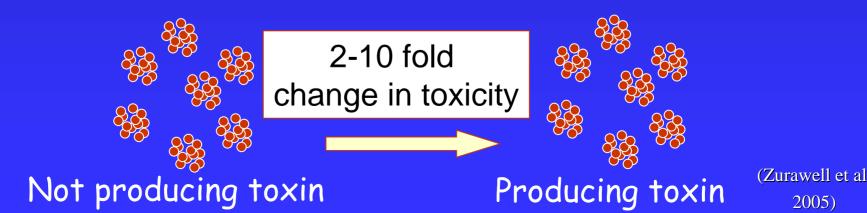


What makes a cyanobacterial bloom toxic?

Shift in community composition



Stimulation of toxin production by environmental factors



2005)

Current projects

Map microcystin concentrations and Microcystis cell numbers in Saginaw Bay and western Lake Erie

Identify environmental factors promoting microcystin production

Develop rapid methods for detection of toxic Microcystis

Accumulation in fish

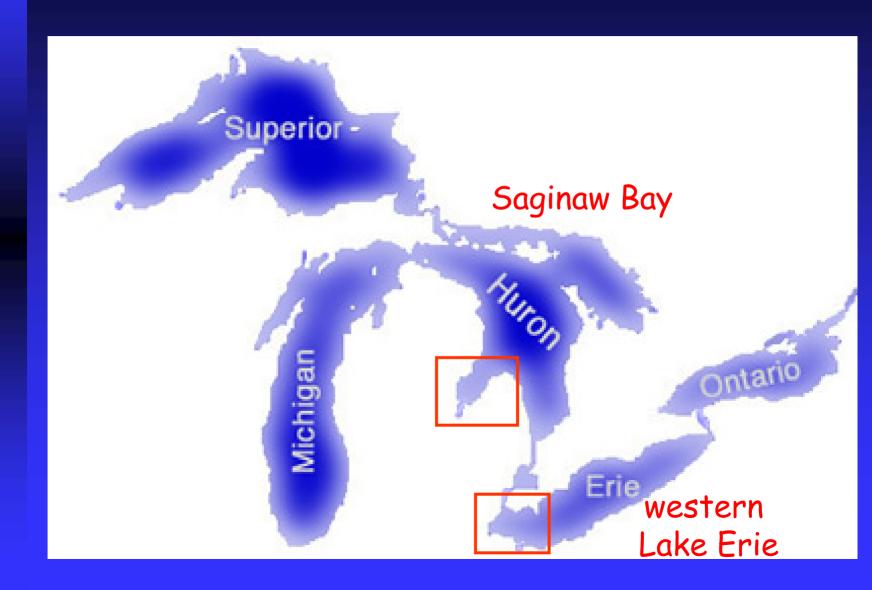
Goal

Develop predictive capabilities for presence of toxic cyanobacterial blooms in Great Lakes recreational and drinking water supplies

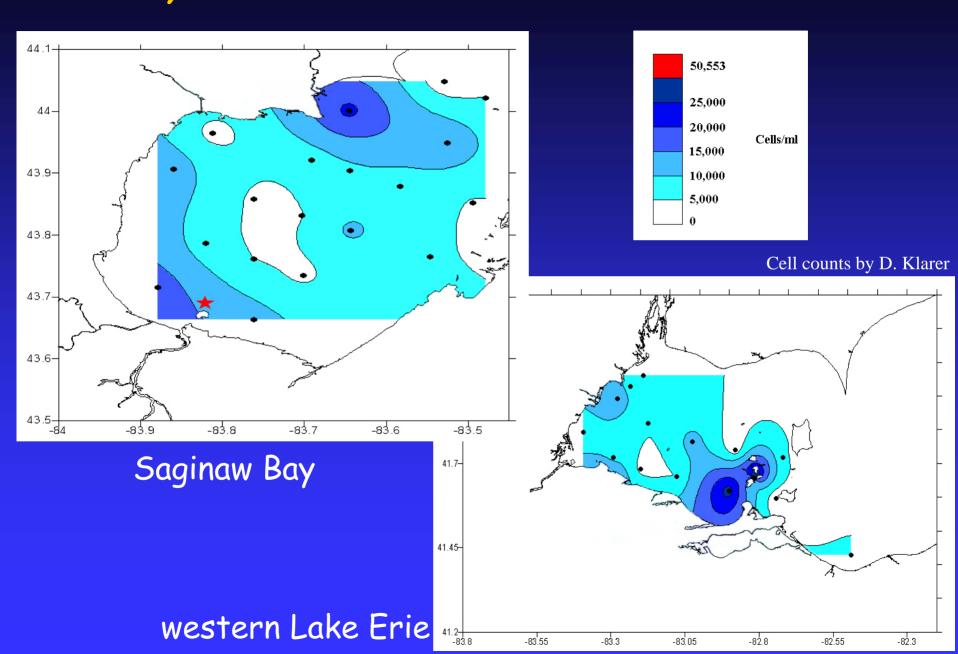




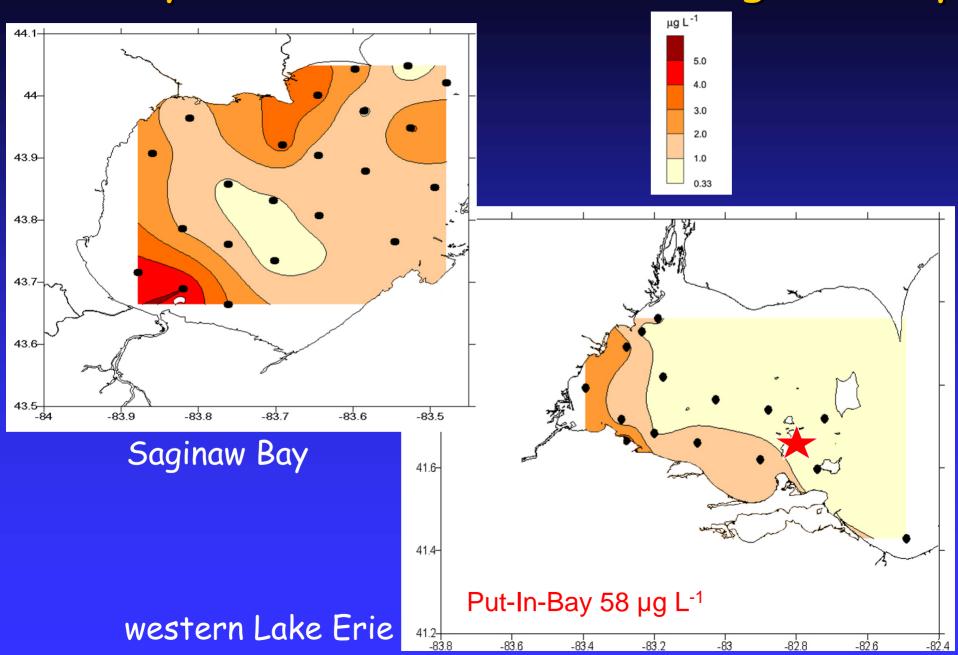
Sampling sites



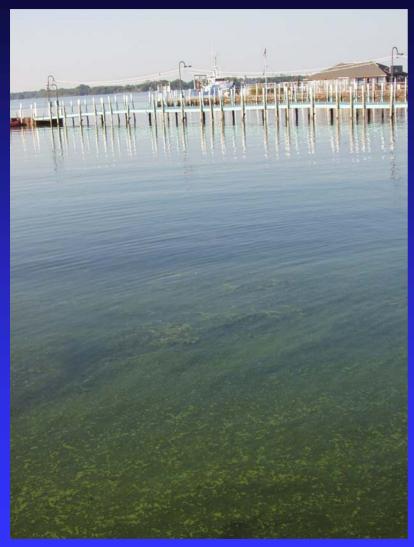
Microcystis cell abundance



Microcystin concentrations in Saginaw Bay



Microcystis in Lake Erie

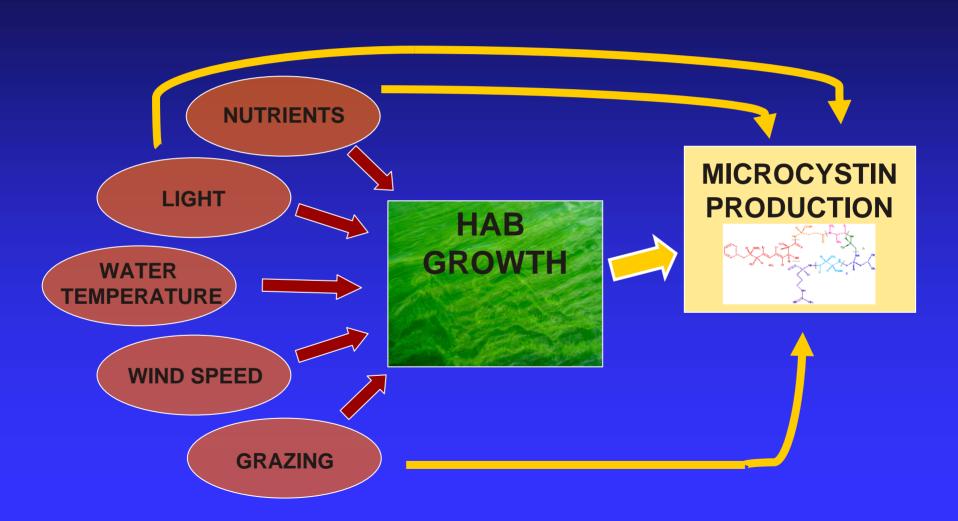


Put-In-Bay
North shore of S. Bass Island



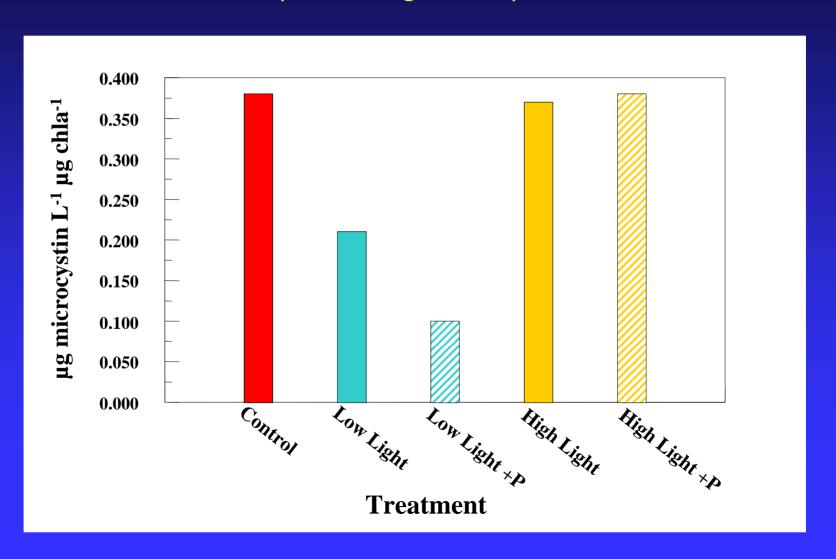
South shore of S. Bass Island

Environmental factors influencing growth and toxin production in *Microcystis*



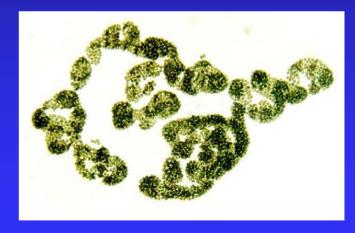
Nutrient (P) and light effects on microcystin production in *Microcystis*-dominated phytoplankton community

Preliminary data, Saginaw Bay, June 2005



Detection with genetic techniques

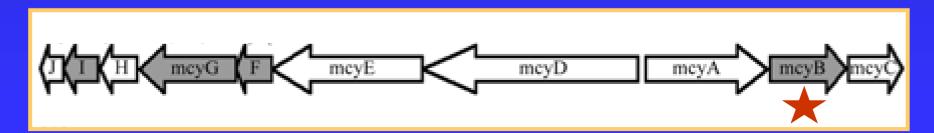
- Differentiate morphologically identical strains
 - toxic vs. non-toxic
- Track specific populations
 - Geographic origin
 - Genetic diversity
- Rapid detection
 - often faster and less tedious than microscopy



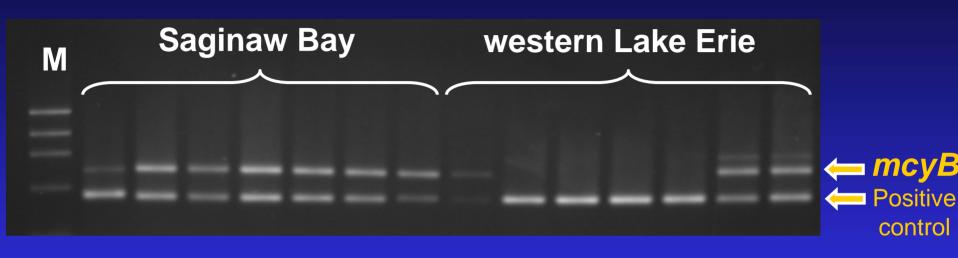
Microcystis sp.

Identifying toxic strains of Microcystis

- All toxin-producing strains of *Microcystis* contain genes for microcystin production: *mcyA-J*
- Presence of mcyB = strain able to produce toxin Absence of mcyB = non-toxic



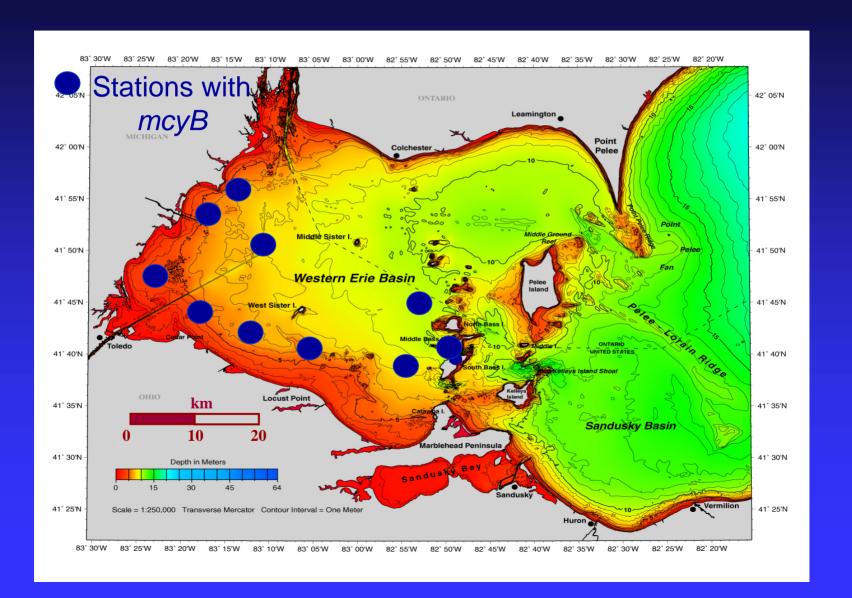
Multiplex PCR for toxic Microcystis



M = molecular weight marker

	Number of colonies		% microcystin
Basin	# mcyB	total	producers
Saginaw	36	40	90%
Erie	4	16	25%

Distribution of toxic Microcystis



Develop a quantitative PCR assay for enumerating toxic Microcystis colonies

Applications

- measure temporal variation in proportion of toxic strains
 - biweekly sampling at 3 locations in western Lake Erie
- identifyir produci
 - zebra
 - chang
- Tie into Microcy



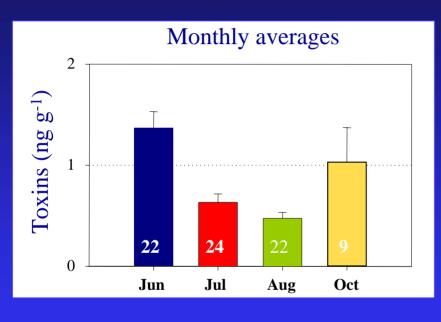
oution of toxic lity

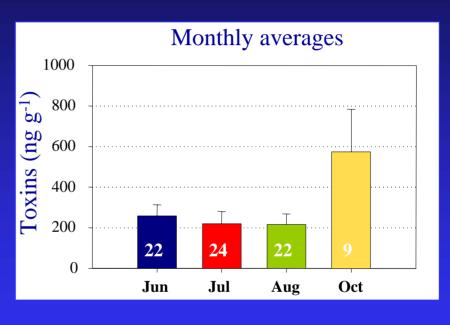
Thanks

- Center of Excellence for Great Lakes and Human Health (Oceans and Human Health Initiative)
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- Hank Vanderploeg (NOAA-GLERL)
- Pat Tester, Wayne Litaker (NOAA-Beaufort)
- Dave Millie (Florida Institute of Oceanography)
- Crew of the R/V Laurentian
- Sabrina Varnam



Microcystin concentrations in Perch Lake Erie, summer 2006 ng toxin (g dry mass)-1





Muscle

Liver

Microcystin concentration of concern for routine fish consumption = 7.7 ng g⁻¹

Microcystis in the Great Lakes

1970

Dominant member of phytoplankton community
Blooms frequent and abundant
High P input to system (detergents, fertilizers, septic)

1980

P abatement programs (Great Lakes Water Quality Agreement)

1990

Decrease in chlorophyll, increased water clarity Blooms rare

Dreissenid mussel introduction

Return of *Microcystis* blooms up to 90% phytoplankton community

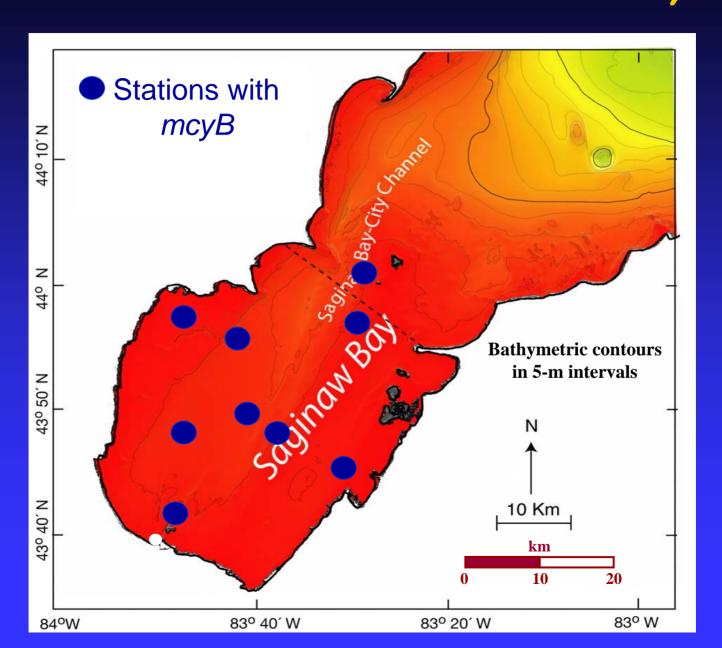
2000

Present

Abundant Microcystis blooms, July - Sept

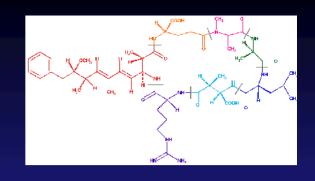


Distribution of toxic Microcystis



Microcystin

- Hepatotoxin
 - Over 60 structural variants



- Health effects
 - Animal mortality: livestock, wildlife, birds, pets
 - Human illness:
 - Gastrointestinal, dermatitis (short term exposure)
 - Liver damage (chronic exposure)
- WHO recommended exposure limits
 - 20 μg/L recreational exposure
 - 1 μg/L drinking water
- Some evidence of bioaccumulation in fish, mussels and zooplankton

